



Full wwPDB NMR Structure Validation Report ⓘ

Mar 9, 2026 – 07:19 AM UTC

PDB ID : 5V16 / pdb_00005v16
BMRB ID : 30261
Title : HnRNP A1 Alters the Conformation of a Conserved Enterovirus IRES Domain
to Stimulate Viral Translation
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Deposited on : 2017-03-01

This is a Full wwPDB NMR Structure Validation Report for a publicly released PDB entry.

We welcome your comments at validation@mail.wwpdb.org

A user guide is available at

<https://www.wwpdb.org/validation/2017/NMRValidationReportHelp>

with specific help available everywhere you see the ⓘ symbol.

The types of validation reports are described at

<http://www.wwpdb.org/validation/2017/FAQs#types>.

The following versions of software and data (see [references ⓘ](#)) were used in the production of this report:

MolProbity : 4-5-2 with Phenix2.0
Percentile statistics : 20250101.v01 (using entries in the PDB archive January 1st 2025)
wwPDB-RCI : v_1n_11_5_13_A (Berjanski et al., 2005)
PANAV : Wang et al. (2010)
wwPDB-ShiftChecker : v1.2
Ideal geometry (proteins) : Engh & Huber (2001)
Ideal geometry (DNA, RNA) : Parkinson et al. (1996)
Validation Pipeline (wwPDB-VP) : 2.49

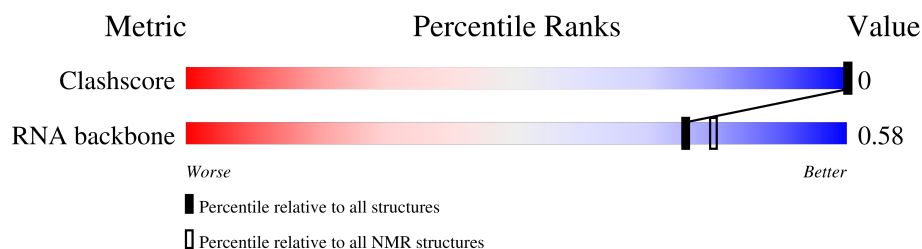
1 Overall quality at a glance

The following experimental techniques were used to determine the structure:

SOLUTION NMR

The overall completeness of chemical shifts assignment is 17%.

Percentile scores (ranging between 0-100) for global validation metrics of the entry are shown in the following graphic. The table shows the number of entries on which the scores are based.



Metric	Whole archive (#Entries)	NMR archive (#Entries)
Clashscore	229148	14424
RNA backbone	8273	777

The table below summarises the geometric issues observed across the polymeric chains and their fit to the experimental data. The red, orange, yellow and green segments indicate the fraction of residues that contain outliers for ≥ 3 , 2, 1 and 0 types of geometric quality criteria. A cyan segment indicates the fraction of residues that are not part of the well-defined cores, and a grey segment represents the fraction of residues that are not modelled. The numeric value for each fraction is indicated below the corresponding segment, with a dot representing fractions $\leq 5\%$

Mol	Chain	Length	Quality of chain
1	A	41	 88% 10% •

2 Ensemble composition and analysis ⓘ

This entry contains 10 models. This entry does not contain polypeptide chains, therefore identification of well-defined residues and clustering analysis are not possible. All residues are included in the validation scores.

3 Entry composition [i](#)

There is only 1 type of molecule in this entry. The entry contains 1313 atoms, of which 444 are hydrogens and 0 are deuteriums.

- Molecule 1 is a RNA chain called RNA (41-MER).

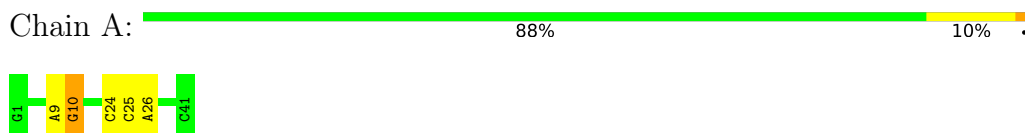
Mol	Chain	Residues	Atoms						Trace
1	A	41	Total	C	H	N	O	P	0
			1313	390	444	156	283	40	

4 Residue-property plots [i](#)

4.1 Average score per residue in the NMR ensemble

These plots are provided for all protein, RNA, DNA and oligosaccharide chains in the entry. The first graphic is the same as shown in the summary in section 1 of this report. The second graphic shows the sequence where residues are colour-coded according to the number of geometric quality criteria for which they contain at least one outlier: green = 0, yellow = 1, orange = 2 and red = 3 or more. Stretches of 2 or more consecutive residues without any outliers are shown as green connectors. Residues which are classified as ill-defined in the NMR ensemble, are shown in cyan with an underline colour-coded according to the previous scheme. Residues which were present in the experimental sample, but not modelled in the final structure are shown in grey.

- Molecule 1: RNA (41-MER)

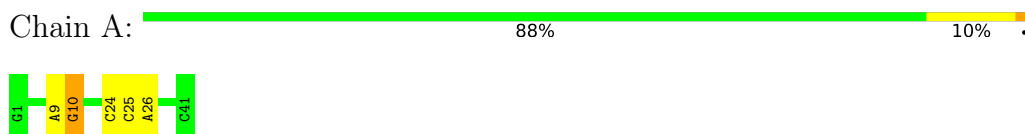


4.2 Scores per residue for each member of the ensemble

Colouring as in section [4.1](#) above.

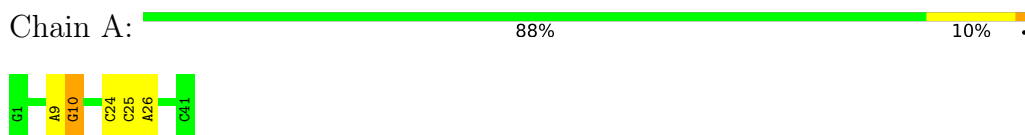
4.2.1 Score per residue for model 1

- Molecule 1: RNA (41-MER)



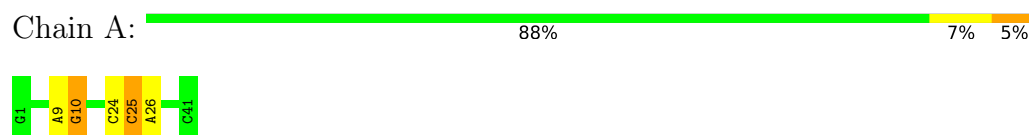
4.2.2 Score per residue for model 2

- Molecule 1: RNA (41-MER)



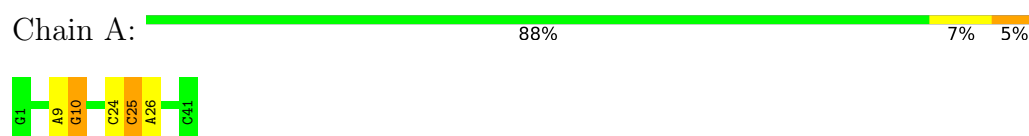
4.2.3 Score per residue for model 3

- Molecule 1: RNA (41-MER)



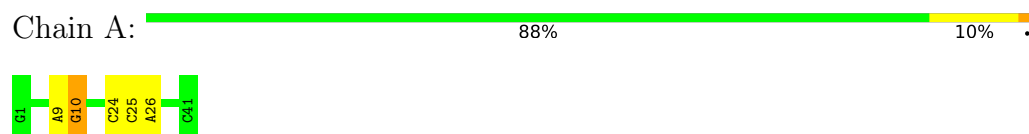
4.2.4 Score per residue for model 4

- Molecule 1: RNA (41-MER)



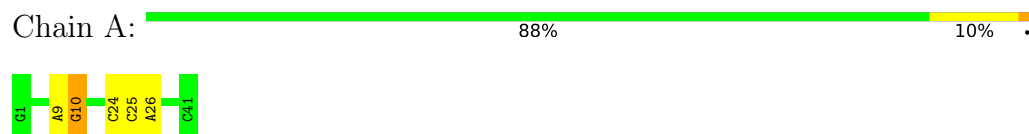
4.2.5 Score per residue for model 5

- Molecule 1: RNA (41-MER)



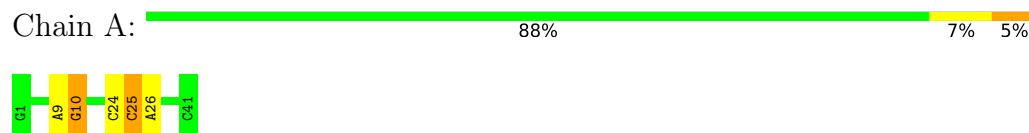
4.2.6 Score per residue for model 6

- Molecule 1: RNA (41-MER)



4.2.7 Score per residue for model 7

- Molecule 1: RNA (41-MER)



4.2.8 Score per residue for model 8

- Molecule 1: RNA (41-MER)

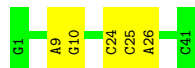
Chain A:  88% 7% 5%



4.2.9 Score per residue for model 9

- Molecule 1: RNA (41-MER)

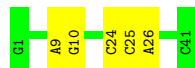
Chain A:  88% 12%



4.2.10 Score per residue for model 10

- Molecule 1: RNA (41-MER)

Chain A:  88% 12%



5 Refinement protocol and experimental data overview

The models were refined using the following method: *molecular dynamics*.

Of the 1024 calculated structures, 10 were deposited, based on the following criterion: *structures with the lowest energy*.

The following table shows the software used for structure solution, optimisation and refinement.

Software name	Classification	Version
X-PLOR NIH	structure calculation	
Amber	refinement	

The following table shows chemical shift validation statistics as aggregates over all chemical shift files. Detailed validation can be found in section 7 of this report.

Chemical shift file(s)	working_cs.cif
Number of chemical shift lists	1
Total number of shifts	131
Number of shifts mapped to atoms	131
Number of unparsed shifts	0
Number of shifts with mapping errors	0
Number of shifts with mapping warnings	0
Assignment completeness (well-defined parts)	17%

6 Model quality [i](#)

6.1 Standard geometry [i](#)

The Z score for a bond length (or angle) is the number of standard deviations the observed value is removed from the expected value. A bond length (or angle) with $|Z| > 5$ is considered an outlier worth inspection. RMSZ is the (average) root-mean-square of all Z scores of the bond lengths (or angles).

Mol	Chain	Bond lengths		Bond angles	
		RMSZ	#Z>5	RMSZ	#Z>5
1	A	0.82±0.00	0±0/971 (0.0± 0.0%)	1.01±0.01	1±1/1511 (0.1± 0.0%)
All	All	0.82	0/9710 (0.0%)	1.01	12/15110 (0.1%)

There are no bond-length outliers.

All unique angle outliers are listed below. They are sorted according to the Z-score of the worst occurrence in the ensemble.

Mol	Chain	Res	Type	Atoms	Z	Observed(°)	Ideal(°)	Models	
								Worst	Total
1	A	10	G	C1'-O4'-C4'	-6.37	103.53	109.90	8	8
1	A	25	C	O4'-C4'-C3'	5.09	109.09	104.00	7	4

There are no chirality outliers.

There are no planarity outliers.

6.2 Too-close contacts [i](#)

In the following table, the Non-H and H(model) columns list the number of non-hydrogen atoms and hydrogen atoms in each chain respectively. The H(added) column lists the number of hydrogen atoms added and optimized by MolProbity. The Clashes column lists the number of clashes averaged over the ensemble.

Mol	Chain	Non-H	H(model)	H(added)	Clashes
All	All	8690	4440	4440	-

The all-atom clashscore is defined as the number of clashes found per 1000 atoms (including hydrogen atoms). The all-atom clashscore for this structure is -.

There are no clashes.

6.3 Torsion angles [i](#)

6.3.1 Protein backbone [i](#)

There are no protein molecules in this entry.

6.3.2 Protein sidechains [i](#)

There are no protein molecules in this entry.

6.3.3 RNA [i](#)

Mol	Chain	Analysed	Backbone Outliers	Pucker Outliers	Suiteness
1	A	40/41 (98%)	4±0 (10±0%)	3±0 (8±0%)	0.64±0.00
All	All	400/410 (98%)	40 (10%)	30 (8%)	0.64

The overall RNA backbone suiteness is 0.58.

All unique RNA backbone outliers are listed below:

Mol	Chain	Res	Type	Models (Total)
1	A	9	A	10
1	A	10	G	10
1	A	25	C	10
1	A	26	A	10

All unique RNA pucker outliers are listed below:

Mol	Chain	Res	Type	Models (Total)
1	A	9	A	10
1	A	24	C	10
1	A	25	C	10

6.4 Non-standard residues in protein, DNA, RNA chains [i](#)

There are no non-standard protein/DNA/RNA residues in this entry.

6.5 Carbohydrates [i](#)

There are no oligosaccharides in this entry.

6.6 Ligand geometry

There are no ligands in this entry.

6.7 Other polymers

There are no such molecules in this entry.

6.8 Polymer linkage issues

There are no chain breaks in this entry.

7 Chemical shift validation [i](#)

The completeness of assignment taking into account all chemical shift lists is 17% for the well-defined parts and 17% for the entire structure.

7.1 Chemical shift list 1

File name: working_cs.cif

Chemical shift list name: *WT_pklist_cln_fn.str3*

7.1.1 Bookkeeping [i](#)

The following table shows the results of parsing the chemical shift list and reports the number of nuclei with statistically unusual chemical shifts.

Total number of shifts	131
Number of shifts mapped to atoms	131
Number of unparsed shifts	0
Number of shifts with mapping errors	0
Number of shifts with mapping warnings	0
Number of shift outliers (ShiftChecker)	0

7.1.2 Chemical shift referencing [i](#)

No chemical shift referencing corrections were calculated (not enough data).

7.1.3 Completeness of resonance assignments [i](#)

The following table shows the completeness of the chemical shift assignments for the well-defined regions of the structure. The overall completeness is 17%, i.e. 131 atoms were assigned a chemical shift out of a possible 771. 0 out of 0 assigned methyl groups (LEU and VAL) were assigned stereospecifically.

	Total	¹ H	¹³ C	¹⁵ N
Sugar	79/451 (18%)	79/246 (32%)	0/205 (0%)	0/0 (—%)
Base	52/320 (16%)	52/197 (26%)	0/72 (0%)	0/51 (0%)
Overall	131/771 (17%)	131/443 (30%)	0/277 (0%)	0/51 (0%)

The following table shows the completeness of the chemical shift assignments for the full structure. The overall completeness is 17%, i.e. 131 atoms were assigned a chemical shift out of a possible 771. 0 out of 0 assigned methyl groups (LEU and VAL) were assigned stereospecifically.

	Total	¹ H	¹³ C	¹⁵ N
Sugar	79/451 (18%)	79/246 (32%)	0/205 (0%)	0/0 (—%)
Base	52/320 (16%)	52/197 (26%)	0/72 (0%)	0/51 (0%)
Overall	131/771 (17%)	131/443 (30%)	0/277 (0%)	0/51 (0%)

7.1.4 Statistically unusual chemical shifts ⓘ

There are no statistically unusual chemical shifts.

7.1.5 Random Coil Index (RCI) plots ⓘ

No *random coil index*(RCI) plot could be generated from the current chemical shift list. RCI is only applicable to proteins